

# Translational Controls in Pain

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28 **Abstract**

29 Pain is an unpleasant but essential sensation. On a cellular level, pain typically originates  
30 in sensory neurons called nociceptors. They undergo rapid increases in cap-dependent  
31 translation in response to noxious stimuli. The specificity of translational controls in  
32 nociceptors is governed by regulatory factors and mRNAs that collaborate to ensure  
33 precise temporal and spatial regulation of protein synthesis. Multiple signaling pathways  
34 bridge extracellular cues to nascent translation including: the mammalian target of  
35 rapamycin (mTOR), AMP-activated protein kinase (AMPK), and the integrated stress  
36 response (ISR). The torrent of information on both mechanisms and targets of  
37 translational controls in nociceptive circuits supports an enticing corollary. Targeted  
38 inhibition of aberrant translation in the cells responsible for the genesis of pain signals in  
39 the periphery affords a new strategy to prevent or reverse chronic pain states. We  
40 describe the implications of emerging insights into translational controls predominantly in  
41 the peripheral nervous system on the search for safer and more specific pain  
42 therapeutics.

43

44 **Keywords:** Pain, ISR, mTOR, AMPK, nociception, translation, anti-nociceptive  
45 mechanisms, hyperalgesia, allodynia

46

## 47 **Introduction**

48

49 The nervous system facilitates a crucial role in detection of harmful cues through a  
50 conserved process termed nociception(Tracey, 2017). It serves a critical function in the  
51 prevention of tissue damage and increases organismal fitness. Humans with congenital  
52 insensitivity to pain (CIP) often perish in childhood due to injuries or infections that fail to  
53 be recognized(Indo et al., 1996). Nociceptors are sensory neurons tasked with detection  
54 of noxious stimuli (e.g. heat, inflammatory cytokines, neurotrophic factors, capsaicin).  
55 They play a key role in both the detection and propagation of pain signals to the spinal  
56 cord that are ultimately communicated to the somatosensory cortex of the brain (**Figure**  
57 **1 A**). After an injury, nociceptors undergo remarkable changes in their activity (termed  
58 plasticity) that often outlive the healing process(Pace et al., 2018). Nociceptor  
59 sensitization refers to a failure of nociceptors to return to their resting state and may play  
60 a major role in the transition from acute to chronic pain(Ferrari et al., 2010). Translational  
61 control have emerged as a dominant theme in nociceptor plasticity(Khoutorsky and Price,  
62 2018, Melemedjian and Khoutorsky, 2015). Here we provide an overview of the  
63 tremendous body of evidence in support of translation as an integral component of pain  
64 signaling. We emphasize the critical role of nociceptors given their key function in the  
65 detection and relay of pain signals.

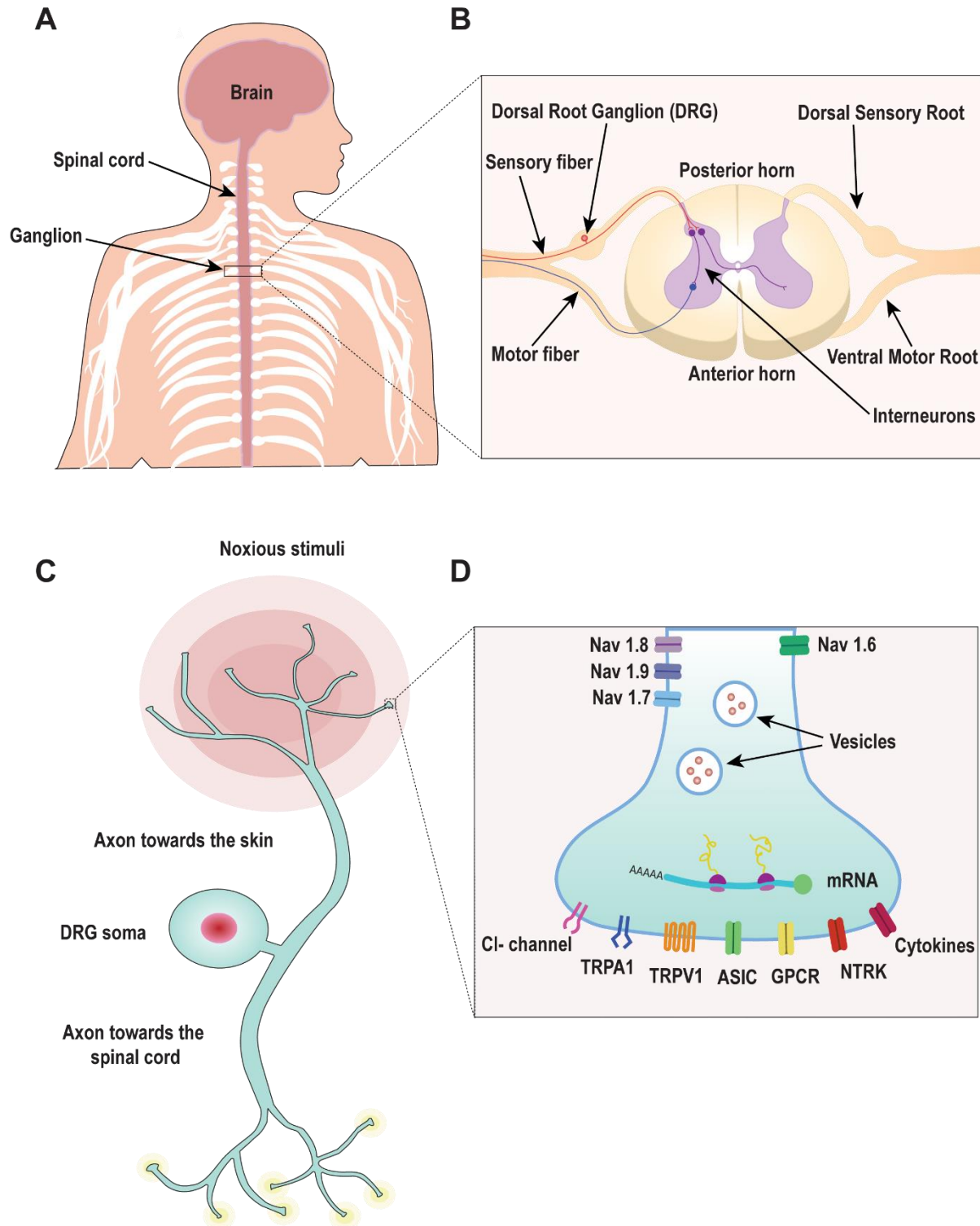
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## 67 **A primer on pain physiology**

68

69 Nociceptors are pseudounipolar neurons tasked with detection of harmful stimuli and  
70 propagation of these signals to the spinal cord (**Figure 1 B**). The nucleus and many of  
71 the organelles in the nociceptor are housed in the cell body or soma. These are found in  
72 two different tissues, the dorsal root ganglia (DRG) adjacent to the spinal cord, or the  
73 trigeminal ganglia (TG) in the head. Approximately half of the neurons in the DRG and  
74 TG are nociceptors in most species. Protein and mRNA expression differ substantially  
75 between cell types giving rise to characteristic conduction velocity, diameter, and stimuli  
76 responsiveness. These differences manifest in the fibers (also known as axons) that  
77 extend from the soma (**Figure 1C**).

78



79

80 **Figure 1** – (A) The anatomy of the pain. (B) Nociceptors are responsible for detecting  
 81 harmful stimuli. Cell bodies of nociceptors are clustered in the DRG adjacent to the spinal  
 82 cord. DRG neurons have one axon with two branches: one branch (sensory fiber)

83 innervates the skin, peripheral tissues, and internal organs. The opposing branch  
84 (sensory root) synapses with neurons in the spinal cord which then relay the signal to the  
85 somatosensory cortex of the brain via the thalamus. (D) Receptors expressed on the  
86 nerve endings of sensory fibers can detect various noxious stimuli such as heat (TRPV1),  
87 environmental irritants (TRPA1), pro-inflammatory mediators (e.g. NGF) (NTRK) and  
88 cytokines. Synaptic vesicles can store neurotransmitters at the synapse and are  
89 controlled by voltage gated calcium channels.

90

91 One end innervates the skin and other peripheral organs, including most of the viscera,  
92 and is responsible for the detection of noxious and/or damaging stimuli.

93

94 Injury can result in damage to the axon and its subsequent degeneration (Davies et al.,  
95 2019). All nociceptor axons are associated with Schwann cells through structures called  
96 remak bundles but most nociceptors are not myelinated. Following injury, Schwann cells  
97 have been demonstrated to play a critical role in the clearance of debris resulting from  
98 tissue degradation and secrete molecules, including nerve growth factor (NGF), that  
99 stimulate axonal growth. This process hinges on local production of proteins in axons  
100 (**Figure 1D**). Axons projecting into the skin shed any associated Schwann cells and  
101 encounter fibroblasts and keratocytes. After an injury, in addition to inflammatory  
102 mediators secreted by immune cells (e.g. IL-6), keratinocytes release ATP which  
103 contributes to changes in nociceptive activity and pain-associated behaviors (Moehring  
104 et al., 2018). Thus, the microenvironment surrounding the nerve fiber facilitates axonal  
105 regeneration and activity.

106

107 The most critical function accomplished by nociceptor axons is signal relay. Peripheral  
108 receptors are electrically silent in a resting state, but once threshold is reached they  
109 transmit action potentials back to the central nervous system (CNS) (Ferrari et al., 2013b,  
110 Ferrari et al., 2013c, Inceoglu et al., 2015, Khoutorsky et al., 2016, Melemedjian et al.,  
111 2010, Xu et al., 2014, Bogen et al., 2012, Moy et al., 2017, Barragan-Iglesias et al., 2018)

112 (Dubin and Patapoutian, 2010). These signals are received by interneurons in the dorsal  
113 horn of the spinal cord. The spinal cord transmits pain signals to the brain, where they  
114 are consciously perceived. Specific neurons act as checkpoints and determine whether  
115 a pain signal is relayed or not, thus not all signals are relayed (Tracey and Mantyh, 2007).  
116 Injury changes the electrophysiological and neurochemistry of neurons that detect and  
117 relay pain signals (Song et al., 2003). Hypersensitivity to noxious stimuli (hyperalgesia)  
118 or innocuous stimuli (allodynia) results from neuronal plasticity that causes a lowering of  
119 pain thresholds (**Figure 2**).

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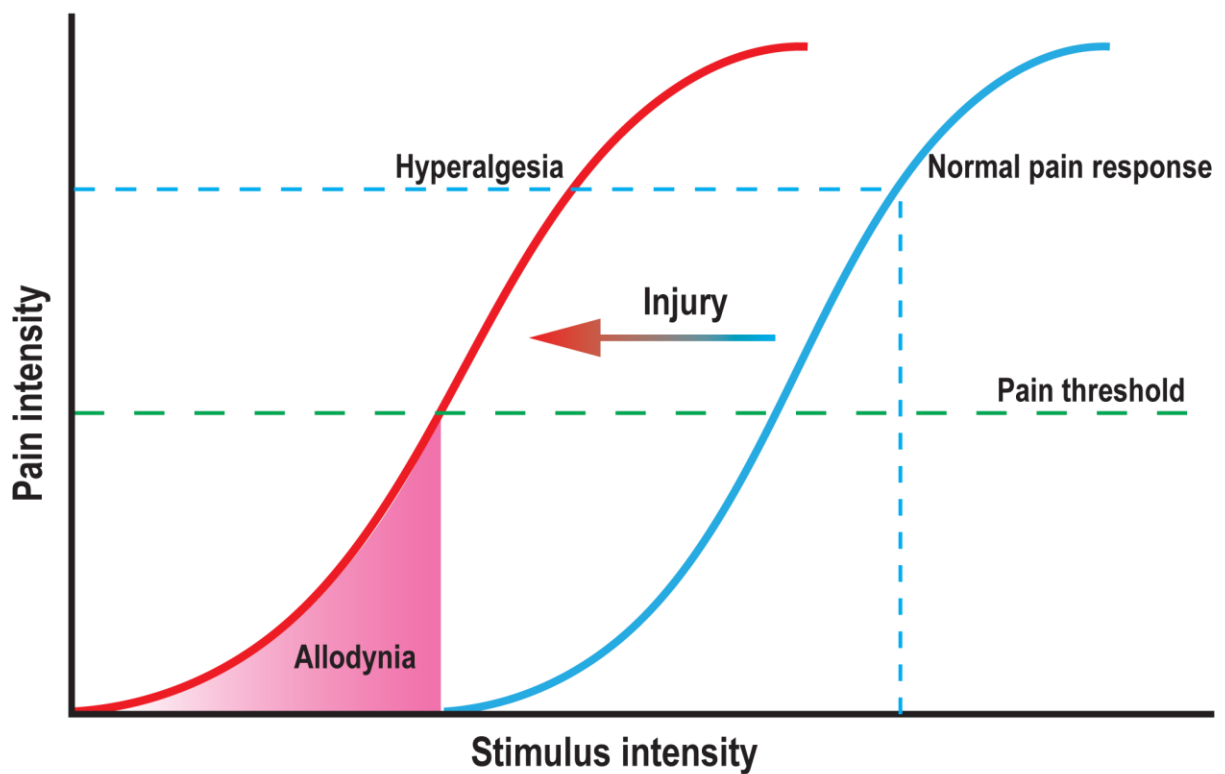
121 A growing body of evidence suggests that translational controls are integral to sustained  
122 changes in neuronal excitability that drive persistent pain states (Megat et al., 2019,  
123 Barragan-Iglesias et al., 2019, Moy et al., 2017, Ferrari et al., 2015a, Ferrari et al., 2015b,  
124 Melemedjian et al., 2014a, Ferrari et al., 2013a, Sonali Uttam, 2018, Khoutorsky et al.,  
125 2016, Khoutorsky et al., 2015, Melemedjian and Khoutorsky, 2015, Melemedjian et al.,  
126 2013a, Melemedjian et al., 2011, Obara et al., 2011, Geranton et al., 2009, Hunt et al.,  
127 2001). A major challenge moving forward is dissecting differential contributions of  
128 translational control to establishment as opposed to maintenance of pain states.  
129 Tremendous insights into nociceptor plasticity have resulted from studies of hyperalgesic  
130 priming. Priming refers to susceptibility to normally subthreshold noxious inputs following  
131 a noxious stimulus. The strength of this model is the ability to separate acute and  
132 prolonged pain states (Kandasamy and Price, 2015). The ever expansive pharmacopoeia  
133 for translational control applied to hyperalgesic priming will expand our understanding of  
134 acute and chronic pain. Increasingly precise genetic and optogenetic tools are also likely  
135 contribute key insights.

136

### 137 **Local translation**

138 Neurons must modulate their function in response to a range of physiologic stimuli. A key  
139 mechanism that facilitates rapid changes in sensory fibers is local translation from  
140 polarized populations of mRNA (Jung et al., 2014). RNA-localization is common in  
141 eukaryotes. An extreme example can be found in *Drosophila* embryos where ~70% of

142 genes show distinct patterns of subcellular localization (Tomancak et al., 2007, Lecuyer  
143 et al., 2007). Most of the corresponding proteins co-localize with their transcripts  
144 suggestive of a potential use of RNA localization to regulate sites of protein synthesis.  
145 Highly specialized cell types, including neurons, make extensive use of RNA localization.  
146 RNA-seq on neuronal processes suggests highly specific mechanisms of mRNA  
147 trafficking (Andreassi et al., 2010, Cajigas et al., 2012, Gummy et al., 2011, Minis et al.,  
148 2014, Zivraj et al., 2010, Taylor et al., 2009). During transit, translation of the mRNA is  
149 repressed often via protein factors recruited to the 3' untranslated region (UTR) (**Figure**  
150 **3A**). The repertoire of RNA-binding proteins bound to mRNAs destined for local



151

152 **Figure 2** – Injury changes pain responses. Allodynia (pain due to a stimulus that does not  
153 usually provoke pain) and hyperalgesia (increased pain from a stimulus that usually elicit  
154 pain) are commonly observed in patients after an injury. Maladaptive central changes and  
155 nociceptor sensitization contribute to the generation and maintenance of allodynia and  
156 hyperalgesia which can ultimately lead to chronic pain (Cervero and Laird, 1996).

157

158 translation is controlled by a variety of signaling mechanisms (Gumy et al., 2011, Taylor  
159 et al., 2009, Zivraj et al., 2010, Willis et al., 2011, Merianda et al., 2009, Yudin et al., 2008,  
160 Willis et al., 2007). These multi-protein complexes serve critical roles in both trafficking of  
161 the mRNA and ensuring that translation is repressed until the transcript has arrived at the  
162 appropriate location within the cell. Thus, tremendous precision is achieved through cis-  
163 acting elements present in mRNA that provide all subsequent regulatory potential by  
164 trans-acting factors.

165

166 Local translation serves a key biological function. Neuronal protein synthesis can occur  
167 in the soma, synapse, or in axons. In nociceptors, axons can span vast distances (in  
168 some cases a meter or longer). Local translation provides a means to accomplish protein  
169 biosynthesis at the site where polypeptides are required. This provides a rapid solution to  
170 the problem of generating new proteins on demand that can guide critical processes to  
171 the function of afferent fibers (such as axonal growth) (Kar et al., 2018, Brittis et al., 2002).  
172 Local translation requires instructions provided by mRNA, and executed through the  
173 combined actions of ribosomes, tRNAs, and regulatory factors. Regulatory factors play a  
174 critical role in triggering translation of the correct target at the appropriate moment when  
175 it is required. Among the best characterized examples of activity-dependent protein  
176 synthesis is local translation of the immediate early gene Arc. Arc is translated in dendrites  
177 as an integral component of learning and memory processes in the hippocampus and  
178 amygdala (Tzingounis and Nicoll, 2006, McIntyre et al., 2005, Guzowski et al., 2000,  
179 Guzowski et al., 1999). However, the role of Arc in peripheral neurons is unclear. While  
180 Arc is translated in the spinal cord, it appears to be dispensable for inflammatory pain  
181 (Hossaini et al., 2010). What are the regulatory features present in mRNA that dictate the  
182 specificity of local translation in nociceptors? While the answer is likely transcript specific,  
183 emerging evidence suggests that analogous mechanisms to neurons in the CNS are  
184 employed, making use of untranslated regions (UTRs) to impart changes in mRNA  
185 localization (Willis et al., 2011, Baj et al., 2016).

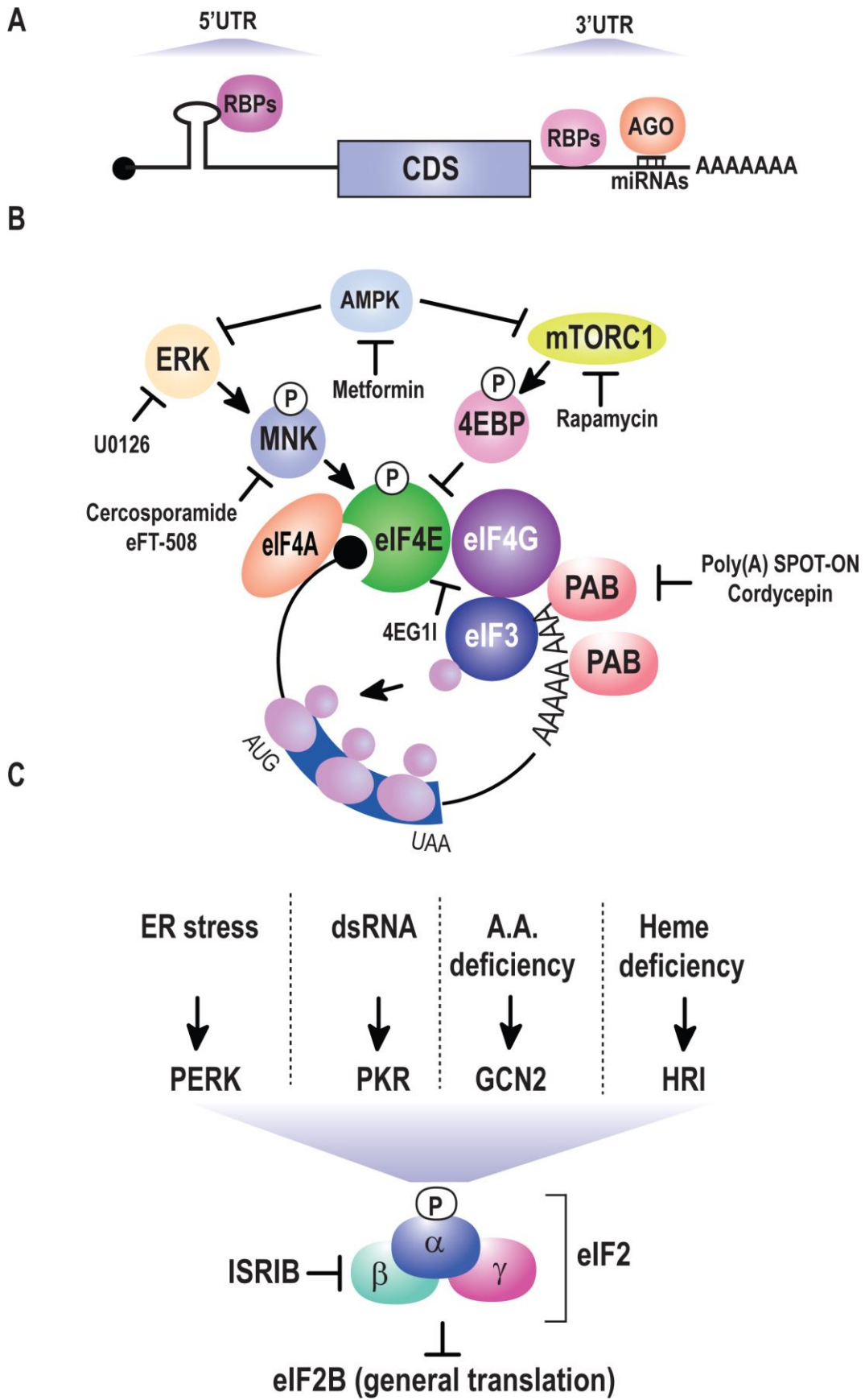
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187 Multiple lines of evidence suggest a key role for local translation in pain. First, axons are  
188 key sites of protein synthesis particularly in nociceptors (Kar et al., 2018, Terenzio et al.,  
189 2018, Merianda and Twiss, 2013, Barragan-Iglesias et al., 2018). Injection of protein  
190 synthesis inhibitors into the paw blocks behavioral responses to inflammatory mediators  
191 that increase nociceptor excitability (Melemedjian et al., 2010, Black et al., 2018). Second,  
192 disruption of mRNA polyadenylation specifically in the DRG blocks hyperalgesic priming  
193 through local translation of CamKII $\alpha$  (Ferrari et al., 2013b, Bogen et al., 2012). Third, NGF  
194 increases axonal localization of a subset of mRNAs (Willis et al., 2005). Fourth, injection  
195 of NGF into humans promotes mechanical hypersensitivity without inflammation through  
196 a mechanism that is locally regulated (Rukwied et al., 2010). Additionally, injection of NGF  
197 into an axonal branch of a single nociceptor sensitizes only that branch (Obreja et al.,  
198 2018). Fifth, proteomics of neuromas suggests that and pulse chase labeling experiments  
199 suggest that local translation of cytoskeletal factors drives hyper-excitability after nerve  
200 damage (Huang et al., 2008). Finally, several groups have identified Nav1.8 mRNA as  
201 axonally localized after peripheral nerve injury (Thakor et al., 2009, Hirai et al., 2017).  
202 Knockdown of Nav1.8 in the sciatic nerve fiber but not the DRG blocks neuropathic pain  
203 caused by sciatic nerve entrapment (Ruangsri et al., 2011).

204

205



207 **Figure 3** – (A) mRNA is comprised of a 5' UTR, the coding sequence (CDS), the 3'UTR,  
208 and the poly(A) tail. The M7G cap structure (black ball) is found on the 5' end of the  
209 transcript. Structures in the 5' UTR can influence translation efficiency and can also recruit  
210 RNA-binding proteins (RBPs). Similarly, the 3' UTR contains regulatory elements that can  
211 be bound by trans-acting RNAs (e.g. miRNAs) as well as proteins. The transcript is  
212 appended with a poly(A) tail. (B) Translation initiation and key inhibitors. AMPK negatively  
213 regulates both mTOR and ERK. Erk controls MNK which ultimately phosphorylates  
214 eIF4E. mTOR controls eIF4E availability through phosphorylation of 4EBPs. eIF4G  
215 interacts with PABP to promote initiation. eIF4G facilitates recruitment of the 40S  
216 ribosomal subunit through interactions with eIF3 (not shown). (C) Stimuli that engage the  
217 ISR are indicated as well as downstream kinases that act on eIF2 $\alpha$ . eIF2 $\alpha$ -dependent  
218 translation is blocked by the small molecule ISRIB.

219

220

221

## 222 mRNA structure

223

224 Eukaryotic mRNAs are resplendent with regulatory features. These include the ubiquitous  
225 5' 7-methylguanosine ( $m^7G$ ) cap on the 5' end of the transcript (**Figure 3A**). The cap is  
226 bound by the cap binding protein eIF4E (Sonenberg et al., 1979). Loss of the  $m^7G$  renders  
227 the mRNA susceptible to rapid 5'→3' degradation by exonucleases (e.g. Xrn1). mRNAs  
228 possess two UTRs that either precede the coding segment on the 5' side or follow the  
229 stop codon on the 3' end. The UTRs harbor regulatory information in the form of *cis*-acting  
230 structures and sequences which are bound by *trans*-acting regulatory factors. These  
231 include RNA-binding proteins and regulatory RNAs that act in consort with RNA-binding  
232 proteins. The 5'UTR has distinct classes of regulatory elements that includes internal  
233 ribosomal entry sequences (IRES) and upstream open reading frames (uORFs). IRES  
234 elements can overcome the need for eIF4E mediated translation initiation through  
235 recruitment of translation factors. The function of uORFs is generally to reduce protein  
236 output of the main reading frame but they can also change the reading frame, add  
237 additional protein sequence, or encode functional peptides (Barbosa et al., 2013). A key

238 property of 5'UTRs is structural content. Secondary structure in the 5'UTR can increase  
239 dependency on the helicase eIF4A and further refine translational output. Similar to the  
240 5' UTR, the 3'UTR can encode binding sites for regulatory factors and serves as a major  
241 repository of information that can enhance or reduce translational efficiency. The 3'UTR  
242 also provides a critical function in neurons as a source of information for specification of  
243 local translation (Menon et al., 2004, Huang et al., 2003, Aronov et al., 2001). Part of the  
244 challenge in elucidating their targets are the dynamic changes in 3'UTR length caused by  
245 alternative polyadenylation (APA). The final step on mRNA maturation is addition of the  
246 Poly(A) tail to the 3' end of the mRNA (AC and M, 2008). Poly(A) tail length is intimately  
247 linked to translational efficacy and the Poly(A) binding protein appears to be integral to  
248 pain signaling(Barragan-Iglesias et al., 2018). APA provides a mechanism to modulate  
249 poly(A) site selection and appears to be critical for localization of ion channels (e.g.  
250 Na<sub>v</sub>1.8) required for nociception (Hirai et al., 2017). Finally, targeted disruption of  
251 polyadenylation by the small molecule cordycepin reverses pain hypersensitivity (Ferrari  
252 et al., 2013a).

253

## 254 **Initiation**

255

256 Protein synthesis is the culmination of a complex process initiated with the birth of RNA  
257 during transcription and the emergence of nascent peptides on the ribosome. Translation  
258 can be described in a series of four subsequent steps – translation initiation, elongation,  
259 termination, and ribosome recycling. Translation initiation is the rate-limiting step and has  
260 garnered tremendous attention as the bulk of translational control is thought to occur at  
261 this step (Hinnebusch et al., 2016). Inhibition of translation initiation in nociceptors  
262 abolishes sensitization and exemplifies the central role that translation initiation plays in  
263 pain plasticity(Melemedjian et al., 2010, Melemedjian et al., 2014b, Moy et al., 2017). In  
264 mammals, the main initiation pathway is termed cap-dependent translation and is  
265 responsible for the initiation of most translational events under non-stress conditions  
266 (Aitken and Lorsch, 2012). However, alternative pathways exist and are essential for  
267 survival under stress conditions and viral infections (Holcik and Sonenberg, 2005).  
268 Among the best-studied examples of alternative initiation pathways are IRES. They reside

269 in the 5'UTR and can directly recruit the ribosome to the mRNA. While their function in  
270 pain is unclear, cellular IRES initiate translation of mRNA subsets when cap-dependent  
271 translation is compromised and could mediate translation of nociceptive factors (Komar  
272 and Hatzoglou, 2011).

273

## 274 **Cap-dependent translation**

275

276 Cap-dependent translation hinges on multiple complexes that recruit the ribosome to the  
277 mRNA (Aitken and Lorsch, 2012) (**Figure 3B**). The eukaryotic initiation factor 4E (eIF4E)  
278 associates with the 5' 7-methylguanosine ( $m^7G$ ) cap of the mRNA (Sonenberg et al.,  
279 1979). eIF4E is controlled both at the level of phosphorylation at a single site and through  
280 sequestration by a protein partner (eIF4E binding protein (4E-BP)) (Pause et al., 1994,  
281 Waskiewicz et al., 1997). eIF4E interacts with the scaffold protein eIF4G which in turn  
282 binds the helicase eIF4A. Collectively, this tripartite complex (referred to as eIF4F) stably  
283 associates with the  $m^7G$  cap. eIF4F phosphorylation globally affects mRNA translation,  
284 and in some cases alters the translation of specific subsets of mRNAs – frequently  
285 proteins that are important for cell survival (Hsieh et al., 2012). Once assembled, eIF4F  
286 recruits the 43S pre-initiation complex (PIC) to the  $m^7G$  cap. The PIC consists of the small  
287 ribosomal subunit (40S) bound to the initiation factor eIF2, initiator tRNA Met-tRNA<sub>i</sub><sup>Met</sup>,  
288 and GTP. Though intrinsically active, eIF4A helicase activity is further stimulated by  
289 complex formation and unwinds the 5' UTR of the mRNA to facilitate ribosomal scanning  
290 of the 5' UTR. Thus, the translation of many mRNAs with highly structured 5' UTRs is  
291 eIF4A-dependent. Upon encountering the AUG start codon, the large ribosomal subunit  
292 (60S) joins the complex to form the 80S ribosome, and eIF2 is released. The joining of  
293 the large ribosomal subunit concludes successful translation initiation and transitions the  
294 ribosome into the elongation phase. eIF4G further interacts with PABP and circularizes  
295 the mRNA, possibly facilitating re-initiation after a successful round of translation (Wells  
296 et al., 1998).

297

## 298 **eIF4F**

299

300 Several signaling cascades converge on eIF4F. Multiple lines of evidence suggest that  
301 eIF4E is central in the development of pain pathologies. While the interaction of eIF4G  
302 with eIF4E is crucial for pain amplification, as evidenced by pharmacological studies (e.g.  
303 4EGI1) (Moerke et al., 2007), a specific role of eIF4A in pain remains poorly understood.  
304 A possible reason might be that the high expression level of eIF4A and its eIF4F-  
305 independent helicase properties complicate tight regulation (Duncan and Hershey, 1983)  
306 (Galicia-Vazquez et al., 2012). In contrast, eIF4E has a low expression level, thus minor  
307 changes in availability by sequestration or modification can have extensive consequences  
308 on translation initiation. Two major pathways directly affect and modulate eIF4E activity;  
309 the mechanistic target of rapamycin (mTOR), and the mitogen-activated protein kinase  
310 (MAPK) pathways (**Figure 3B**) (Melemedjian et al., 2010, Moy et al., 2017).

311  
312 The mechanistic target of rapamycin (mTOR) signaling cascade is a dominant regulatory  
313 feature of translational control (Yanagiya et al., 2012). The mTOR catalytic subunit exists  
314 in two multimeric protein complexes, one of which is sensitive to inhibition by rapamycin  
315 (mTORC1). In neurons, the mTORC1 pathway receives input from a large variety of  
316 upstream pathways that relay external input to mTORC1 which in turn creates cellular  
317 responses (Boutouja et al., 2019). mTORC1 upstream receptors include: NMDA, Trk, and  
318 IGF-1. The downstream targets of mTOR include regulators of translation like eIF4E  
319 binding proteins (4E-BPs), p70 S6 kinase (S6K), and eEF2 kinase. The three known 4E-  
320 BP isoforms (1, 2, and 3) show a tissue specific expression and the predominant isoform  
321 in the pain processing pathway is 4E-BP1 (Jimenez-Diaz et al., 2008, Melemedjian et al.,  
322 2011, Xu et al., 2010, Khoutorsky et al., 2015). Phosphorylation of 4E-BPs releases eIF4E  
323 from sequestration and allows it to engage in the eIF4F complex. Inflammatory pain  
324 models using injections of the upstream activators nerve growth factor (NGF) and  
325 interleukin 6 (IL-6), revealed a rapid induction of protein synthesis in nociceptors, which  
326 is concurrent with the activation of mTORC1 as monitored by phosphorylation  
327 (Melemedjian et al., 2010). Conversely, pharmacological inhibition of mTORC1 with  
328 rapamycin-related small molecules reduces pain hypersensitivity in a wide variety of pain  
329 models (Geranton et al., 2009, Jimenez-Diaz et al., 2008, Price et al., 2007). The  
330 endogenous endothelial growth factor receptor (EGFR) ligand, Epregrulin (EREG),

331 stimulates the mTOR pathway in DRG neurons and upregulates matrix metalloproteinase  
332 9 (MMP-9) translation (Martin et al., 2017). MMP-9 is a regulator of inflammation and is  
333 transiently upregulated in DRG sensory neurons in models of neuropathic chronic pain  
334 (Manicone and McGuire, 2008, Kawasaki et al., 2008). Inhibitors of EGFR, used in cancer  
335 treatments, have been reported to also alleviate pain in patients with cancer-induced  
336 neuropathic pain (Kersten and Cameron, 2012, Moryl et al., 2006).

337  
338 While mTORC1 is a global regulator of translation, it also appears to alter translation  
339 locally in the sciatic nerve and proprioceptive DRG neurons. During neuronal injury mTOR  
340 is transiently activated and translation of its own mRNA and other survival promoting  
341 molecules is up-regulated in a 3' UTR-dependent fashion (Terenzio et al., 2018). 3' UTRs  
342 frequently contain localization motifs suggesting that local mRNA pools can be deposited  
343 and activated upon a stimulus, in this case injury. Local pharmacological repression of  
344 mTOR leads to reduced neuron numbers. It is not known, however, if injury-induced local  
345 translation of mTOR affects nociception plasticity.

346  
347 mTORC1 is also known to specifically regulate specific subsets of transcripts. For  
348 example, mTOR regulates expression of mRNAs that contain terminal oligopyrimidine  
349 tracts in their 5' UTRs (5' TOP mRNAs) via 4E-BPs (Thoreen et al., 2012). A critical issue  
350 in the field is systematic identification of mTOR targets that contribute to pain associated  
351 behaviors. While the molecular mechanisms by which these subsets are selected remains  
352 elusive an enticing hypothesis is that disabling cap-dependent translation favors  
353 alternative initiation pathways. While so far not investigated in nociceptors, this  
354 hypothesis is underpinned by increased IRES-dependent translation of Arc mRNA in  
355 dendrites when cap-dependent initiation is inhibited (Pinkstaff et al., 2001), which is  
356 consistent with the continued translation of IRES-containing mRNAs in the presence of  
357 mTOR inhibitors (Torin-1) (Thoreen et al., 2012).

358  
359 S6K1 and 2 are downstream effectors of mTORC1. S6Ks that act on translation  
360 elongation by phosphorylation of initiation and elongation factors like eukaryotic  
361 elongation factor 2 (eEF2) (reviewed in (Zoncu et al., 2011)). Although an important

362 regulator of elongation, the role of S6K1/2 in pain is less clear than that of eIF4E. The  
363 investigation of S6K1 has been complicated by predominantly relying on genetic tools as  
364 small molecules targeting S6Ks lack in high specificity. In models of chronic inflammation  
365 pain, mTOR activation leads to S6K1 phosphorylation in DRG neurons but remains  
366 unaffected in neuropathic pain models (Liang et al., 2013). S6K1/2 double-knockout mice  
367 are more sensitive to mechanical stimuli with unaltered thermal sensitivity. The direct  
368 implications of S6K1/2 on elongation are masked by a negative feedback mechanism that  
369 in the long term activates the MAPK/ERK pathway. This leads to hyperexcitability of  
370 sensory neurons, allodynia, and spontaneous pain (Melemedjian et al., 2013a), which  
371 makes S6K1/2 a poor target for pharmacological intervention.

372  
373 The MAPK pathway controls phosphorylation of a single residue, Ser209, in eIF4E via  
374 MAPK-interacting protein kinases (MNKs) 1 and 2 (Pyronnet et al., 1999, Waskiewicz et  
375 al., 1999). MNK1/2-mediated eIF4E phosphorylation contributes to the development of  
376 nociceptor sensitization and promotes chronic pain after injury (Moy et al., 2017). Both,  
377 phosphorylation-resistant eIF4E<sup>S209A</sup> mutant mice, and reciprocally, MNK knockout mice  
378 show decreased pain hypersensitivity in response to most inflammatory mediators.  
379 Inhibition of eIF4E phosphorylation also inhibits hyperalgesic priming (Melemedjian et al.,  
380 2010, Moy et al., 2017). Similar to the mTOR pathway, MNK1/2-dependent  
381 phosphorylation of eIF4E Ser209 is suspected to promote tissue-specific alternative  
382 translation of mRNA subsets. Few eIF4E-phosphorylation dependent mRNA targets have  
383 been identified so far. In the pain processing pathway, known targets are matrix  
384 metalloproteases (MMP-2 and 9) and the key regulator of pain plasticity, *Bdnf*, in dorsal  
385 root ganglia (Moy et al., 2018). Translation of *Bdnf* mRNA is stimulated in response to  
386 inflammation and is important for pain plasticity and hyperalgesia (Obata and Noguchi,  
387 2006) (Melemedjian et al., 2013b, Moy et al., 2018, Melemedjian et al., 2014a). In the  
388 DRG, eIF4E phosphorylation is required for hyperalgesic priming and promotes the  
389 translation of a specific *Bdnf* mRNA isoform (Bdnf-201), which has the longest and most  
390 structured 5' UTR of all *Bdnf* isoforms (Moy et al., 2018). The specific translation  
391 enhancement might reflect the stimulatory role of eIF4E on the RNA helicase eIF4A,  
392 though the specific effect of eIF4E-phosphorylation is unknown. These findings highlight



393 that local and tissue specific eIF4E-dependent translation is a feature of pain  
394 amplification.

395

### 396 **eIF4E in chemotherapy induced peripheral neuropathy**

397

398 While several relevant pathways for pain amplification have been identified, the  
399 translational alterations of their specific mRNA targets are mostly unknown. A ribosome  
400 profiling study identified regulators of the MAPK/ERK pathway as mRNA targets in the  
401 DRG and spinal cord dorsal horn in neuropathic pain (Sonali Uttam, 2018). Though  
402 ribosome profiling lends itself to the identification of translationally regulated mRNA  
403 expression, the cellular heterogeneity of the nervous system poses an obstacle and can  
404 confound the identification of cell type-specific changes in protein expression. A method  
405 that allows for cell-type specific analysis is translating ribosome affinity purification  
406 (TRAP) (Heiman et al., 2008), which uses a tagged ribosomal protein that is specifically  
407 expressed in the desired cell type. An initial study using TRAP has described translation  
408 in nociceptors in chemotherapy (paclitaxel)-induced pain in mice (Megat et al., 2019).  
409 Sequencing of mRNA bound to tagged ribosomes and further pharmacological and  
410 mutational validation suggests that MNK1 mediated eIF4E phosphorylation increases  
411 translation of the mTORC1-activator RagA complex. In mice, pain-associated behavioral  
412 effects of paclitaxel were reversed upon injection of a MNK inhibitor called eFT508. This  
413 suggests that pharmacological disruption of cap-dependent translation may provide a  
414 means to reverse neuropathic pain states. Consistent with this notion, elimination of the  
415 sole phosphorylation site on eIF4E results in profound deficits in pain behavioral  
416 responses to inflammatory mediators (Moy et al., 2018, Moy et al., 2017). This work  
417 suggests that cap-dependent translation is integral to the persistence chemotherapy-  
418 induced neuropathic pain.

419

### 420 **eIF2**

421

422 eIF2 is another key regulator of protein translation that promotes initiation (Holcik and  
423 Sonenberg, 2005), and is a known effector in neuropathic pain (Barragan-Iglesias et al.,

424 2019). Phosphorylation on Ser51 of the eIF2 $\alpha$  subunit is the nexus of four pathways that  
425 collectively form the integrated stress response (ISR) (Khoutorsky et al., 2016, Sidrauski  
426 et al., 2015). These pathways (**Figure 3C**) are activated by viral infection (double-  
427 stranded RNA-dependent protein kinase, PKR), ER-stress (PKR-like ER kinase, PERK),  
428 amino acid deprivation (general control non-repressible 2, GCN2), oxidative stress and  
429 heme-deficiency (heme-regulated inhibitor, HRI) (Lu et al., 2001). Ser51 inhibits initiation  
430 by turning eIF2 into a competitive inhibitor of its GDP exchange factor (GEF) eIF2B,  
431 rendering it inactive (Yang and Hinnebusch, 1996, Pavitt et al., 1998, Krishnamoorthy et  
432 al., 2001, Jennings et al., 2013). eIF2 $\alpha$  phosphorylation is increased in models of diabetes  
433 induced neuropathic pain and chronic inflammation (Barragan-Iglesias et al., 2019)  
434 (Khoutorsky et al., 2016). The targetability of individual pathways and subsequently the  
435 phosphorylation state of eIF2 $\alpha$  make it an attractive pharmacological target. For example,  
436 activation of eIF2B by the small molecule ISRIB (Tsai et al., 2018) reverts eIF2 $\alpha$   
437 phosphorylation via PERK and relieves both translational inhibition and diabetic pain in  
438 mice (Barragan-Iglesias et al., 2019).

439  
440 eIF2 $\alpha$  phosphorylation generally inhibits translation but stimulates translation of upstream  
441 ORFs (uORFs) in the 5' UTRs of mRNAs (Barbosa et al., 2013, Hinnebusch et al., 2016).  
442 eIF2 $\alpha$  phosphorylation can also impact read-through of uORFs through eIF2A-dependent  
443 mechanisms (Sendoel et al., 2017). Thus, it is tempting to speculate that this transient  
444 shift from main ORFs (mORF) to uORFs causes pain hypersensitivity, potentially by  
445 affecting the local biophysics of the cell and membrane. The molecular mechanism by  
446 which uORFs affect nociception remains to be investigated.

447

#### 448 **eIF2 $\alpha$ in diabetic peripheral neuropathy**

449

450 A reactive glycolytic metabolite associated with painful diabetic pain called methylglyoxal  
451 triggers neuropathic pain via the integrated stress response (Barragan-Iglesias et al.,  
452 2019). Intriguingly, MGO induced pain or diabetic pain caused by ablation of insulin  
453 producing cells (with streptozotocin) is reversed by the small molecule inhibitor ISRIB

454 that targets eIF2B. While the relevant targets are unknown, the integrated stress  
455 response has been broadly implicated in neuronal function and is likely key in a variety of  
456 pain states linked to increases in eIF2a phosphorylation. Indeed, hemizygous loss of  
457 eIF2a phosphorylation decreases thermal but not mechanical hypersensitivity  
458 (Khoutorsky et al., 2016).

459

## 460 **AMPK**

461

462 AMP-activated protein kinase (AMPK) functions as a key energy sensor and has emerged  
463 as a therapeutic target for pain (Price et al., 2015, Taylor et al., 2013, Price and Dussor,  
464 2013, Carling et al., 2012). Three subunits contribute to AMPK function ( $\alpha$ ,  $\beta$ , and  $\gamma$ ). The  
465  $\gamma$  subunit senses the AMP/ATP ratio and mediates allosteric effects on the  $\alpha$  subunit. The  
466 catalytic domain is modulated by an upstream kinase (AMPKK). AMPK controls mTOR  
467 via two different pathways. AMPK directly inhibits mTOR activity through phosphorylation  
468 of raptor and indirectly inhibits mTOR via activation of the TSC complex. AMPK is a target  
469 for metabolic disease and cancer. AMPK agonists including metformin attenuate nascent  
470 translation and increase neuronal p-granules (Melemedjian et al., 2014a). AMPK  
471 activators appear to attenuate allodynia caused by peripheral nerve injury and reduce the  
472 excitability of nociceptors *in vitro* (Melemedjian et al., 2011).

473

## 474 **Translational controls in the central nervous system**

475

### 476 **Re-consolidation mechanisms in pain**

477

478 Reconsolidation has been coupled to protein synthesis inhibitors as a means of erasing  
479 memories and has clear implications for traumatic memories that can lead to pathological  
480 states (e.g. posttraumatic stress disorder). Analogous states may underlie certain  
481 nociceptive pain states. For example, mechanical hyperalgesia can be labile and  
482 susceptible to reversal by intrathecal delivery of protein synthesis inhibitors (Bonin and  
483 De Koninck, 2014). This work suggests that pain reconsolidation is likely spinally  
484 mediated and could be a useful strategy to reverse persistent pain.

485

486 **Spinal modulation**

487

488 Injury can increase the excitability of nociceptors and of the spinal cord circuitry. Central  
489 sensitization refers to increases in the excitability of the spinal circuit and play a major  
490 role in pain signaling. Central sensitization can amplify signals originating in the periphery  
491 (communicated by the nociceptors) destined for processing by the central nervous  
492 system. The implications are manifest in three ways: allodynia, hyperalgesia, and  
493 generalized pain to noninjured sites (secondary hyperalgesia). Central sensitization is  
494 driven in part by changes in synaptic strengthening at the dorsal horn.

495

496 A major structural model for understanding plasticity comes from understanding the key  
497 role of synaptic strength in learning and memory in the brain. Synaptic strength is  
498 modulated by opposing processes termed long-term potentiation (LTP) and long-term  
499 depression (LTD) in mammals. Learning and memory and LTP share several  
500 commonalities. Both LTP and long-term memory require protein synthesis and are  
501 blocked by mTOR inhibitors (Costa-Mattioli et al., 2009, Martin et al., 1999). Drugs that  
502 block LTP induction also attenuate hyperalgesia *in vivo* (Ruscheweyh et al., 2011).  
503 Finally, electrical stimulation that induces LTP in rodents generates long-lasting increases  
504 in pain perception in humans (Biurrun Manresa et al., 2018). Conversely, electrical  
505 devices that induce LTD show some promise in reduction of pain perception and may be  
506 useful for treating chronic pain (Rottmann et al., 2010).

507

508 **Opioid-induced hyperalgesia**

509

510 Chronic administration of opioids can sensitize patients to acute pain through an effect  
511 called opioid induced hyperalgesia (OIH). Intriguingly, mTOR is activated in the dorsal  
512 horn of the spinal cord in a model of OIH (Xu et al., 2014). This drives an increase in  
513 nascent protein synthesis and eIF4E activity due to an increase in 4E-BP1  
514 phosphorylation. OIH induced mechanical hyperalgesia can be reversed by intrathecal  
515 delivery of rapamycin. While the precise site of action is unclear as the delivery route is

516 not specific to the spinal cord, these data suggest that neuroplasticity in the nervous  
517 system caused by opioids is controlled, at least in part, by mTOR signaling.

518

519

## 520 **Conclusions**

521 Tremendous human suffering results from poorly managed pain. Chronic pain is  
522 estimated to impact the lives of a quarter of the population in the United States  
523 (Dahlhamer et al., 2018). Existing therapies for the treatment of chronic pain include  
524 numerous opioids that interact with reward circuitry in the central nervous system  
525 contributing to their rampant misuse(Pathan and Williams, 2012). Advances in  
526 understanding the genesis of pain particularly in the peripheral nervous system have  
527 tremendous potential value in the identification of new therapeutic targets. Therapeutics  
528 with a peripheral site of action may provide safe and effective alternatives to opiates  
529 because they need not cross the blood brain barrier and target pain from where it  
530 originates. Translational control in peripheral sensory nociceptors has emerged as  
531 important regulator in pain sensitization and in the development and maintenance of  
532 various chronic pain conditions. Despite the identification of upstream signaling events  
533 that mediate translation in nociceptors, we still haven't elucidated the precise mechanism  
534 by which translation leads to nociceptor hyperexcitability and synaptic plasticity. Which  
535 mRNAs are efficiently translated or repressed during a particular pain state, and what are  
536 their functions? What are common among these mRNAs? Can these factors be targeted  
537 for inhibition? Hopefully, the answers to these key questions will provide the genesis for  
538 more effective pain management.

539

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541

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